



Department
for Environment
Food & Rural Affairs



FIELD GUIDE TO INVASIVE ALIEN PLANT PESTS IN THE CARIBBEAN UK OVERSEAS TERRITORIES



PART 6 – INVERTEBRATES (Except insects) and References

Chris Malumphy, Sharon Reid, Rachel Down, Jackie Dunn
and Debbie Collins

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Frontispiece

Top row: Giant African land snail *Lissachatina fulica* © C. Malumphy; Mediterranean fruit fly *Ceratitidis capitata* © Crown copyright; Sri Lankan weevil, *Myloccerus undecimpustulatus undatus* adult © Gary R. McClellan. Second row: Cactus moth *Cactoblastis cactorum* caterpillar © C. Malumphy; Cottony cushion scale *Icerya purcashi* © Crown copyright; Red palm mite *Raoiella indica* adults © USDA. Third row: Tomato potato psyllid *Bactericera cockerelli* © Fera; Cotton bollworm *Helicoverpa armigera* © Crown copyright; Croton scale *Phalacrocooccus howertoni* © C. Malumphy. Bottom row: Red palm weevil *Rhynchophorus ferrugineus* © Fera; Tobacco whitefly *Bemisia tabaci* © Crown copyright; Brown marmorated stink bug *Halyomorpha halys* © David R. Lance, USDA APHIS PPQ, Bugwood.org.

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6.48 Giant African Land Snail

Class: Gastropoda
 Order: Stylommatophora
 Family: Achatinidae
 Species: *Lissachatina fulica* (Bowdich)

	Present	Threat		
	Absent	Bio	Hlth	Econ
Ang	I			
Ber	i	ü	ü	ü
BVI	i	ü	ü	ü
Cay	i	ü	ü	ü
Mon	i	ü	ü	ü
TCI	i	ü	ü	ü



Figure 6.48.1 Giant African Land Snails *Lissachatina fulica*, Mauritius © C. Malumphy

Background

Giant African land snail *Lissachatina fulica* is a major plant pest, native to East Africa, that has been introduced accidentally to many parts of the world with trade and/or intentionally as a food source, for scientific research and education, and as a novelty pet (CABI, 2017; Global Invasive Species Database, 2017). It is readily transportable over considerable distances as it can go into a state of aestivation in cooler or unfavourable conditions. It has established in many tropical regions where it has had a major negative impact due to its broad polyphagy and prodigious reproductive capacity. As a result, *L. fulica* has been classified as one of the world's top 100 invasive species by The World Conservation Union, IUCN (ISSG, 2003). The snail can be found in agricultural areas, coastland, natural forest, planted forests, riparian zones, shrublands, urban areas, and wetlands.

Lissachatina fulica presents a potentially serious plant health threat to all the UKOTs with tropical climates, particularly to those located in the Caribbean where the snail is currently expanding its geographical range.

Geographical Distribution

Lissachatina fulica is native to the coastal regions of East Africa but has become widely distributed by the activities of man in humid tropical areas of sub-Saharan Africa, the Indian Ocean, Asia, Pacific, South America and parts of the Caribbean (Anguilla, Barbados, Guadeloupe, Martinique and Saint Lucia) (Fig. 6.48.2). Incursions in Florida, USA, are under eradication (CABI, 2017; Global Invasive Species Database, 2017).



Figure 6.48.2 *Lissachatina fulica* distribution map © CABI

Host Plants

Lissachatina fulica is broadly polyphagous feeding on hundreds of plant species (Raut & Ghose, 1984). In its native range its preferred food is decayed vegetation and animal matter, lichens, algae and fungi. It was only recognised as a major plant pest after having been introduced into new environments and has since been recorded feeding on many plants including ornamentals, vegetable and fruit crops. It shows a preference for brassicas, breadfruit (*Artocarpus altilis*), cassava (*Manihot esculenta*), cocoa (*Theobroma cacao*), papaya (*Carica papaya*), peanut (*Arachis hypogaea*), rubber (*Hevea brasiliensis*) and most species of legumes and cucurbits. It can also feed on the bark of relatively large trees such as citrus, papaya, rubber and cacao. The snail needs a source of calcium to build its shell and is recorded eating sand, very small stones, bones from carcasses and even concrete (CABI, 2017; Global Invasive Species Database, 2017).

Description

Mature *L. fulica* are distinctive in appearance and are readily identified by their large size and relatively long, narrow, conical shell (Figs 6.48.1, 6.48.3-4 and 6.48.6-8). The shell is usually in the size range 5 to 10 cm but may reach a length of 20 cm or more. The colour is variable but is most commonly light brown, with alternating brown and cream bands on young snails and the upper whorls of larger specimens. The coloration becomes lighter towards the tip of the shell, which is almost white. There are from six to nine spirally striate whorls with moderately impressed sutures. The shell aperture is ovate-lunate to round-lunate with a sharp, un-reflected outer lip (Fig. 6.48.8). The mantle is dark brown with rubbery skin. There are two pairs of tentacles on the head: a short lower pair and a large upper pair with round eyes situated at the apex. The mouth has a horned mandible containing some 80,000 teeth. Eggs (Fig. 6.48.5) are spherical to ellipsoidal in shape (4.5-5.5 mm diameter) and are cream to yellow in colour.

There are several species of large land snail distributed across sub-Saharan Africa, though three are most frequently encountered: *L. fulica* from East Africa is somewhat smaller than its similar counterpart in West Africa, *Achatina marginata*, and has a more pointed apex to its shell. Another similar species, *Achatina achatina*, also from West Africa, is the world's largest snail with a maximum recorded shell length of 27 cm and a weight of almost 1 kg. *Lissachatina fulica* could also be confused with Giant South American Snails (*Megalobulimus* spp.) which are also invasive in parts of the Caribbean. *Megalobulimus* shells are elongate-oval, apex conical, thick, calcareous and generally cream coloured. Mature *Megalobulimus* shells are generally smaller than those of *L. fulica* and usually lack the brown and cream banding found in the latter species (CABI, 2017; Global Invasive Species Database, 2017).



Figure 6.48.3 *Lissachatina fulica* fully extended body searching for food, Brazil © C. Malumphy



Figure 6.48.4 *Lissachatina fulica*, ribbon-like excrement is indicated by the arrow, Singapore © C. Malumphy



Figure 6.48.5 *Lissachatina fulica* eggs hidden under a stone in a National Park, Saint Lucia © C. Malumphy



Figure 6.48.6 Group of *Lissachatina fulica* snails in a small park in an urban area, Saint Lucia © C. Malumphy



Figure 6.48.7 *Lissachatina fulica* shell, dorsal view © Fera



Figure 6.48.8 *Lissachatina fulica* shell, ventral view © Fera

Biology

Lissachatina fulica is hermaphroditic and after a single mating can produce five or six batches of fertile eggs a year. Each batch contains 100 to 400 eggs (average around 200), with up to 1200 being laid in a year. They are laid on the ground, often at the base of plants. These hatch after about 8-21 days under tropical conditions. The reproductive rate of *L. fulica* is remarkable. For example, 20

tonnes of snails were collected on one day in Fiji just 4 years after its introduction. Adults have an average life span of 5-6 years but may live up to 9 years. Although *L. fulica* is a tropical snail, it can survive cold conditions, even snow, by aestivating for up to three years. It is normally nocturnal and crepuscular in its habits, though it will become active in the daytime during rainy or overcast periods (CABI, 2017; Global Invasive Species Database, 2017).

Dispersal and Detection

The snails can move over short distances between host plants (Fig. 6.48.3) but long-distance dispersal is due to anthropogenic activities. They have been moved internationally accidentally with trade and intentionally as a commercial food source, for scientific research, educational purposes, and as a novelty pet. All developmental stages may become attached to plant and non-plant materials, such as agricultural machinery and vehicles. Eggs and juvenile snails are also readily transported in garden waste.

Mature snails are large and conspicuous, and hide during the day, coming out at dusk to feed (Fig. 6.48.1). Surveys are therefore best carried out at night using a flashlight. Mature snails are easily detected, and attacked plants exhibit extensive rasping and defoliation. The sheer weight of numbers can break the stems of some host plant. Its presence can also be detected by signs of ribbon-like excrement (Fig. 6.48.4), and large slime trails on plants and buildings.

The eggs and juvenile snails can be difficult to detect during phytosanitary inspections of imported plants and other cargo.

Economic and other Impacts

Lissachatina fulica can have an economic, environmental and social impact when introduced to new areas (CABI, 2017; Global Invasive Species Database, 2017). The snail has a voracious appetite and has been recorded attacking a wide range of crops. It is considered by most authorities to be the most damaging land snail in the world. Indirectly, *L. fulica* may damage plants by vectoring plant diseases, for example, it has been implicated in the transmission of the fungus *Phytophthora palmivora*.

Snail populations can reach very high densities and invade native ecosystems. It can therefore pose a serious environmental and conservation threat by eating native plants, modifying habitats, and out-competing native species of snail.

High densities of snails in urban areas can be a major nuisance and have a significant social impact. Not only are they unsightly but cadavers smell and make a mess, especially where they are run over by traffic, which invariably happens during population explosions. *Lissachatina fulica* can act as a vector of the human disease, eosinophilic meningitis, which is caused by the rat lungworm parasite, *Angiostrongylus cantonensis*. The parasite is passed to humans through eating raw or improperly cooked snails. In sub-Saharan Africa, *L. fulica* is an important source of protein and large numbers of snails are regularly imported into the UK for human consumption. They can also be used to make fertilizer, chicken feed and fish feed. The snails are used for scientific research and educational purposes, and in Europe and parts of North America (where it is lawful), they are kept as a novelty pet.

6.49 Bumblebee Millipede

Class: Diplopoda
 Order: Spirobolida
 Family: Rhinocricidae
 Species: *Anadenobolus monilicornis* (von Porat)

	Present	Threat		
	Absent	Bio	Hlth	Econ
Ang	i		ü	ü
Ber	i		ü	ü
BVI	i		ü	ü
Cay	l		ü	ü
Mon	i		ü	ü
TCI	l		ü	ü



Figure 6.49.1 Bumblebee millipedes *Anadenobolus monilicornis* collected at a tourist resort in the Turks and Caicos Islands (TCI) © C. Malumphy

Background

Anadenobolus monilicornis is a millipede native to the southern Antilles that is now widespread in the Caribbean and has been introduced to South and North America (Shelly, 2014). It has several common names including yellow-banded millipede, bumblebee millipede or Jamaican bumblebee millipede. Populations can explode after rain and it can become a nuisance when hundreds of millipedes invade houses and tourist resorts. If threatened it has the potential to release a toxic irritant that can cause a dark chemical burn to the skin (Fig. 6.49.7). It is not a primary plant pest but can have a social impact. It has the potential to establish in all the UKOTs in the Caribbean. Shelly (2014) has published a detailed and comprehensive review of *A. monilicornis* and much of the information presented in this factsheet comes from this source.

Geographical Distribution

Anadenobolus monilicornis is native to the southern Antilles, but has been spread by anthropogenic activities in the Caribbean, South Eastern USA and Bermuda. In the Caribbean it has been recorded from Antigua, Barbados, Cayman Islands, Dominica, Dominican Republic, Grenada, Grenadines, Guadeloupe, Haiti, Jamaica, Saint Kitts, Saint Lucia, Saint Vincent, Trinidad and Tobago, Saint Maarten, Martinique, Puerto Rico and the US Virgin Islands.



Figure 6.49.2 Adult bumblebee millipede *Anadenobolus monilicornis* © Totodu74

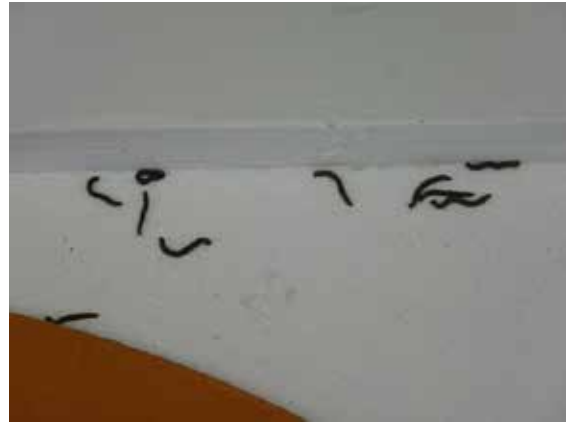


Figure 6.49.3 *Anadenobolus monilicornis* in a tourist resort in TCI © C. Malumphy



Figure 6.49.4 Cleaning away *Anadenobolus monilicornis* in the TCI © C. Malumphy



Figure 6.49.5 *Anadenobolus monilicornis* invading buildings in the TCI © C. Malumphy



Figure 6.49.6 *Anadenobolus monilicornis* curled up in defensive position © Totodu74



Figure 6.49.7 Skin damage caused by toxins from *Anadenobolus monilicornis* © Marc Manfredi

Host Plants

Anadenobolus monilicornis feeds primarily on decaying plant material, as well as higher-quality foods, such as fallen fruits, seeds, mushrooms, faeces and dead invertebrates. It feeds opportunistically on whichever is most available at the soil surface. It does not normally feed on healthy plants but is commonly found in mulch used to fertilize garden plants.

Description

Anadenobolus monilicornis are elongate, cylindrical, with many body segments, most bearing two pairs of legs; they are dark brown to black with distinctive yellow bands, and measure 2.5–10 cm in length (Figs 6.49.1-2 and 6.49.6). The legs and antennae are red. Young millipedes resemble the adults.

Biology

This species inhabits leaf litter and eggs are laid in soil or moist organic matter. Birds and captive monkeys have been observed crushing these millipedes and rubbing their secretions on their wings or fur, probably to repel insects. This millipede might sometimes crawl on an unwary person at night while sleeping. If it finds itself pressed up against the skin, or crushed, it will release a toxic irritant that will cause a dark chemical burn to the skin (Fig. 6.49.7). These darkened areas may take more than 4 months to completely return to normal.

Dispersal and Detection

Anadenobolus monilicornis have a limited natural dispersal potential and can only crawl over relatively short distances but they may be dispersed over long distances with plant trade, for example, eggs and immatures are easily overlooked when hidden in the soil of potted plants. The adults are relatively easy to detect due to their conspicuous colour pattern and hundreds of millipedes may be found entering buildings after rain.

Economic and other Impacts

Anadenobolus monilicornis is not a primary plant pest but is often associated with urban gardens. They can have an economic and social impact as staff in tourist complexes may have to sweep up the millipedes from outside and inside buildings because there is a risk that tourists could receive a chemical burn from the millipedes (Figs 6.49.3-5). They often enter buildings and houses through wall holes, around service lines, around loose-fitting windows or doors, or through roof vents. Once in a building they usually dry out and die relatively quickly unless they find a humid area such as a shower or bathroom. They may also emit an odour when disturbed.

6.50 Red Palm Mite

Class: Arachnida
 Order: Trombidiformes
 Sub order: Prostigmata
 Family: Tenuipalpidae
 Species: *Raoiella indica* (Hirst)

	Present	Threat		
	Absent	Bio	Hlth	Econ
Ang	i			ü
Ber	i			ü
BVI	i			ü
Cay	l			ü
Mon	i			ü
TCI	l			ü



Figure 6.50.1 Red palm mite *Raoiella indica* adults © USDA

Background

Red palm mite, *Raoiella indica*, is a pest of several important ornamental and fruit-producing palm species in the Middle East and South-East Asia. It has invaded the Western Hemisphere and is in the process of colonizing islands in the Caribbean, as well as other areas on the mainland. The invasion of this species is the biggest mite explosion ever observed in the Americas.

Geographical Distribution

The red palm mite originated in India, Egypt, Israel, Mauritius, Reunion, Sudan, Iran, Oman, Pakistan and the United Arab Emirates. *Raoiella indica* was first reported in the New World in Martinique (Flechtmann & Étienne, 2004) and has since spread rapidly throughout the Caribbean archipelago into Southern Florida and South America, and has now spread further into Mexico, Brazil and Colombia (CABI, 2017). It has become of interest as an invasive in these countries due to the high population numbers and diverse range of host plants that the mite has been recorded on. It is spreading rapidly and is expected to establish in other subtropical regions of the Western Hemisphere.

Host Plants

The red palm mite is a pest of several ornamental and fruit-producing palm species such as coconut (*Cocos nucifera*) and areca palms (*Areca* spp.); it has also been found attacking bananas (*Musa* spp.) and plantains (*Musa paradisiaca*) in the Caribbean. Ornamental palms, including hurricane or princess palm (*Dictyosperma album*) and Christmas palm (*Veitchia merrillii*) have also been damaged. Palms are important components of our tropical landscapes, both indoors and out, and in many countries coconut palms (*C. nucifera*) and date palms (*Phoenix dactylifera*) are important food crops.



Figure 6.50.2 A colony of *Raoiella indica* on a coconut leaf © Bryony Taylor



Figure 6.50.3 Close-up of the different life stages of *Raoiella indica* © SENASICA



Figure 6.50.4 Male and female adult *Raoiella indica* © USDA

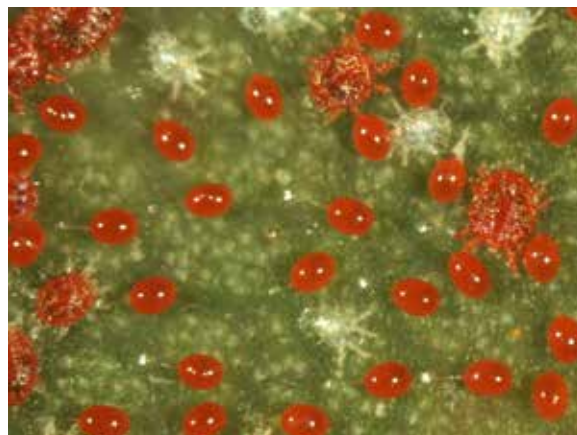


Figure 6.50.5 *Raoiella indica* eggs with immatures and moulted skins (left) and adults (right) © USDA

The red palm mite has been collected from ornamental plants other than palms such as *Heliconia* spp., bird of paradise (*Strelitzia* spp.) and gingers (*Zingiber* spp.) in the Caribbean islands where it has invaded, but it is not clear whether these are valid host plants or whether the enormous mite populations on coconuts and other palms have temporarily moved on to sub-story plants under the palms.

Description

Raoiella indica is a tiny red mite, which is characterized by the presence of long spatulate setae on its dorsum, often with a drop of liquid on the end (Fig. 6.50.1 and 6.50.4). The body shape is oval and flattened and the male can be distinguished from the female by the distinct triangular abdomen (Kane & Ochoa, 2006; Welbourn, 2006). All stages of the mite are red; however, the adult females often have darkened areas on their abdomen (Fig. 6.50.4). There are five distinct life stages: egg, larva, protonymph, deutonymph and adult (Figs 6.50.3-5). The length of the adult female is between 267-300 μm and the width between 178 and 215 μm (Sadana, 1997). The eggs (Fig. 6.50.5) are laid in groups and are approximately 117 μm long, red/orange and smooth and shiny in appearance, and are found attached to the leaf by a stipe that is roughly twice as long as the egg (Kane & Ochoa, 2006).

Biology

Mites in the family Tenuipalpidae are commonly called "false spider mites" and are all plant feeders. However, only a few species of tenuipalps are of economic importance. The tenuipalps have stylet-like mouthparts (a stylophore) similar to those possessed by spider mites (Tetranychidae). The mouthparts are long, U-shaped, with whiplike chelicerae that are used for piercing plant tissues. Tenuipalps feed by inserting their chelicerae into plant tissue and removing the cell contents. These mites are small and flat, and usually feed on the under surface of leaves. They are slow moving and do not produce silk, unlike many of the tetranychid (spider mite) species, which do produce silk.

The number of eggs laid varies from individual to individual; however, on average, 28.1 eggs are laid during the average adult female life span of 27 days. As the larvae and nymphs pass through each stage, they enter a quiescent stage for 36-48 hours, whereby they enter ecdysis and withdraw posteriorly from the exuviae (Zaher *et al.*, 1969). The duration of each stage on coconut at 24.2°C in Mauritius was egg: 4-6 days; larva: 6-8 days; protonymph: 4-7 days; deutonymph: 4-5 days; however, the duration of stages increases with lower average temperatures (Moutia, 1958).

Dispersal and Detection

This mite is easily distributed by wind currents, which are the most likely method of natural dispersal, for example, the presence of mites on older palms on islands adjacent to Martinique within a year of introduction indicate that wind dispersal was the primary dispersal method. However, it is believed that *R. indica* was first accidentally introduced to the Caribbean on infested material imported via shipping lanes from the Old World. No research to date has identified whether this was a single or multiple introduction. Accidental spread to new areas is via the introduction of infested plants or plant material or on palm handicrafts infested with mites or eggs, which are common tourist souvenirs and readily transported. Quarantine measures are in place to prevent the transfer of mites via palm handicrafts, cut flower and leaf arrangements from host plants, and coconut seed.

Inspection of the underside of leaflets of host plants using a hand lens, or removal of material and inspection under a dissecting microscope can confirm the presence of mites. Colonies often contain mites of all stages as well as exuvial remains (white cast skins) and can have up to 300 individuals. Affected host plants will generally display symptoms under heavy infestations. Typical damage symptoms include localized yellowing of the leaf, which can spread into larger chlorotic patches. Heavy infestations may be found along the midrib of coconut leaflets and damage may progress from localized yellowing to necrosis. Infestations on banana and plantain often cause yellowing along the margins of the leaf. If there are heavy infestations, persons inspecting the host plant may find they pick up red on their fingers from the mites on the underside of leaves.

Economic and other Impacts

Raoiella indica is expected to cause economic damage to tropical and subtropical agriculture and to urban and indoor environments where palms are grown. Palm nurseries, landscape palms, and horticultural gardens could also be affected. Coconut growers have reported a 70% drop in coconut production since the introduction of the mite and FAO figures have shown a reduction in coconut production from Caribbean countries since 2004, when the mite was first identified in the region, to the present date. Empirical studies are required to confirm these figures/correlations; however, officials are concerned that a reduction in coconut production may lead to job losses and major socio-economic problems. In Florida, it was feared possible that economic impacts could come from quarantine restrictions if *R. indica* was detected in palm nurseries.

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- Zaher, M. A., Wafa, A. K. & Yousef, A. A. 1969. Biological studies on *Raoiella indica* Hirst and *Phyllotranychus aegyptiacus* Sayed infesting date palms in U.A.R. (Acarina-Tenuipalpidae). *Zeitschrift fur Angewandte Entomologie*, **63** (4): 406-411.
- Zimmermann, H., Bloem, S. & Klein, H. 2004. Biología, Historia, Amenaza, Monitoreo y Control de la Palomilla del Nopal, *Cactoblastis cactorum*. Joint FAO/ International Atomic Energy Agency Programme Nuclear Techniques in Food and Agriculture. Mexico. 61 pp.

9. Appendices

9.1 Major sources of further information

The following list of websites provide a wealth of freely accessible information on invasive alien species that pose a potential plant health risk to the UKOTs in the Caribbean. They also provide links to further sources of information:

CABI. 2007. Crop Protection Compendium. <http://www.cabicompendium.org/>.

CABI. 2017. Invasive species Compendium. <http://www.cabi.org/isc>.

CABI. 2018. Plantwise. <https://www.plantwise.org/>

EPPO. 2018. European and Mediterranean Plant Protection Organization (EPPO) <https://www.eppo.int/>

FDACS 2018. Florida Department of Agriculture and Consumer Services <https://www.freshfromflorida.com/>

NNSS. 2018. GB Non-Native Species Secretariat – UK Overseas Territories Home – <http://www.nonnativespecies.org/ots/otsMap.cfm>

ISSG. 2014. Global Invasive Species Database. <http://www.iucngisd.org/gisd/>

9.2 Fera invertebrate plant pest identification Services for the UKOTs

Defra provide funding to Fera to deliver an identification service for suspected invasive alien invertebrate plant pests in the UKOTs. A submission form and instructions on how to submit a sample are given below.

<h2>UK Overseas Territories Sample Submission Form</h2> <p>To help us with our diagnosis, please try and fill in as much information as possible?</p>	
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For Fera use only:

Your Details

Name:		Date:	
Department/Organisation:		Your Ref:	
Address:		Tel:	<input type="checkbox"/> *
		Fax:	<input type="checkbox"/> *
		Email:	<input type="checkbox"/> *

* Tick preferred method of contact

The Sample

Type of sample:	Plant/Produce Import Interception <input type="checkbox"/> Locally grown plant(s) <input type="checkbox"/> Other (give details) <input type="checkbox"/>		
Type of pest:	Insect <input type="checkbox"/> Mite <input type="checkbox"/> Nematode <input type="checkbox"/> Other (give details) <input type="checkbox"/> Unknown <input type="checkbox"/>		
What do you suspect the pest to be?			
Approximate numbers of organisms found:		Life-stage(s) (if known):	
Sample preservation: (see over for recommended methods)	Live <input type="checkbox"/> Dried/pinned <input type="checkbox"/> Slide mounted <input type="checkbox"/> Fluid preserved (please give details) <input type="checkbox"/>		

The Host

Plant genus, species:	
Description of host damage (if any):	
Country of origin:	
Other information:	

I hereby authorise Fera to carry out diagnostic work on this sample.

Signature:

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Sending your sample for diagnosis

How to collect, package and send your sample

How a sample is preserved before posting is largely dependant on how long it will take to reach us. Please contact us for advice if you are unsure of how to collect and package a particular sample or if you wish to send photos of pests and/or pest damage before submitting a sample. Please email: ukot@fera.co.uk

Collecting and Packaging:

- Provide as many individual specimens of each suspected species as possible. Some species are highly variable, or there may be mixed populations of related species on a host.
- In some cases, adult life stages are necessary for species determination. Larval forms of most Diptera (flies) and Lepidoptera (butterflies and moths) should ideally be collected with their food source and reared on before sending.

Beetles, adult Lepidoptera, and other robust bodied invertebrates	Kill before sending, freezing is preferable. Carefully place the dead insect in a small tube or box and lightly wrap it with tissue (enough to stop it rattling around and becoming damaged). Avoid airtight containers due to possible mould formation. Prevent this by puncturing a few holes in the tube or box. Moths and butterflies can also be placed in paper or glassine envelopes, but take care not to crush them. Pinning, carding and setting of organisms before posting is welcomed, please contact us if you require instructions.
Larval invertebrates	Place in a small plastic tube filled with ethanol (70%).
Scale insects, aphids, whitefly pupae and mites	<p>Leave on the host where possible. Removing them can damage them making identification more difficult. Place the host leaf or stem in a plastic pot of 70% ethanol. With larger leaves or stems, or if the sample is going to take a while to reach us it is necessary to change the preservative after 4-5 days as the plant material will dilute the ethanol and the invertebrates will begin to rot.</p> <p>If you have to detach specimens from the host material this is best done with a fine paintbrush or fine forceps. Place them in a small plastic container filled with ethanol (70%). Do not send specimens in formalin or any other formaldehyde based preservative.</p>
Soil samples	For nematode analysis. Place about 500g of soil in a strong plastic bag and seal.
Sticky traps	Label, bend round sticky side inwards, and secure in position with adhesive tape.

Labelling and Posting:

- Each sample must be clearly labelled indicating host association, date collected, locality, reference numbers and any other pertinent information. Pictures of associated host damage are also useful.
- All samples should be put in a strong cardboard box and packaged securely. Include your Sample Submission Form licence and permits seal the box. We will provide you with a copy of our licence 'Letter of Authority'.
- On the outside of the parcel a label should state "Dead Invertebrates for Scientific Study - No Commercial Value". Please also place a copy of the appropriate permits and licences in an envelope and secure to the outside of the parcel.
- If the parcel is being sent by courier, please send us the tracking reference if possible.

Send your sample to:

UKOT Project (Room 02FA04)
Fera Science Ltd
Sand Hutton
York
YO41 1LZ
United Kingdom

If you have any queries, contact:

Sharon Reid or Dr. Chris Malumphy
Tel: +44 (0)1904 462000
E-mail: ukot@fera.co.uk